

EFFICACY OF SOME ACARICIDES AGAINST ERIOPHYID MITE, *ACERIA* SP. (ACARINA: ERIOPHYIDAE) IN NORTH WEST HIMALAYAS

SANJAY KUMAR SHARMA^{1*} AND PREM CHAND SHARMA²

¹CSKHPKV, Highland Agricultural Research and Extension Centre, Kukumseri, Lahaul and Spiti -175142, Himachal Pradesh, INDIA

²Department of Entomology, CSKHPKV Palampur-176062, Himachal Pradesh, INDIA

e-mail: sanjayhpau@yahoo.com

KEYWORDS

Acaricides
Aceria sp.
Blister development
Eriophyid population
Hippophae

Received on :
29.01.2016

Accepted on :
24.02.2016

***Corresponding author**

ABSTRACT

Field experiments were conducted to evaluate efficacy of four acaricides namely dicofol 18.5EC (0.5 ml/L), propargite 57EC (0.2 ml/L), fenpyroximate 5EC (2.0 ml/L) and hexythiazox 5.45EC (1.8 ml/L) against eriophyid mite, *Aceria* sp. (Acarina: Eriophyidae) infesting seabuckthorn (*Hippophae* L.) in North West Himalayas. The results revealed that two sprays of these acaricides done at ten days interval reduced mite population by 98.8-99.4, 99.5-99.6, 96.1-99.3 and 84.1-99.5 per cent over pre-treatment as observed 10 days after second spray (DAS₂) in two years respectively. Whereas, mite population in control plants increased by 233.5-246.8 per cent over pre-treatment during the same time interval. Likewise, reductions in blister count on seabuckthorn leaves in two years varied from 63.2-78.6, 66.7-81.0, 68.8-70.0 and 56.3-70.0 per cent over pre-treatment as observed at 10DAS₂ in case of dicofol 18.5EC (0.5 ml/L), propargite 57EC (0.2 ml/L), fenpyroximate 5EC (2.0 ml/L) and hexythiazox 5.45EC (1.8 ml/L) sprayed plants, respectively. While, leaves on the control plants registered 100-105.9 per cent increase in blister count during this same time. From present studies, it was concluded that the evaluated acaricides effectively checked the population build-up of eriophyid mite, prevented blister development and could be used for management of eriophyid menace on seabuckthorn.

INTRODUCTION

An eriophyid mite, *Aceria* sp. (Acarina: Eriophyidae) successfully thrives on seabuckthorn (*Hippophae* L.) (Sharma *et al.*, 2011) that exists over an area spanning approximately 12000 ha (Singh, 2003) in highlands of North West Himalayas. It is an immensely important but utterly underutilized plant that grows naturally in these high altitude regions and bears orange yellow berries in bunches. The real worth of this plant lies in rich treasure of industrially useful compounds found in its different parts. For instance, seabuckthorn berries have flavonols such as rutin, quercetin, myricetin, kaempferol, isorhamnetin (Hibasami *et al.*, 2005), tocopherols, tocotrienols (Kallio *et al.*, 2004; Luhua *et al.*, 2004) and carotenoids (Pintea *et al.*, 2001, Weller and Breithaupt, 2003) which possess antioxidant properties (Chaman *et al.*, 2011) and are useful in pharmaceutical formulations intended for improving the human immune system. Its seeds are rich in vitamin C, carotenoids, vitamin E, flavonoids, kaempferol, fatty acids, triacylglycerol, phytosterols, sugar, organic acids, proanthocyanidins and phenolic compounds (Abid *et al.*, 2007; Fan *et al.*, 2007; Li *et al.*, 2007). Besides this, bark and leaves of this plant also have many useful compounds such as flavonoids, carotenes, volatile oils, carbohydrates, vitamins, amino- and mineral acids that are important in pharmaceutical, nutraceutical and cosmetic industries (Catherine *et al.*, 2012; Sabir *et al.*, 2005; Wang *et al.*, 2011; Zeb, 2004a). Various other extracts of this plant also exhibit marked antioxidant

activity (Chauhan *et al.*, 2007; Dhyani *et al.*, 2011; Geetha *et al.*, 2002; Negi *et al.*, 2005; Singh *et al.*, 2011; Suleyman *et al.*, 2002). The slender and worm like eriophyid mites possess piercing and sucking mouthparts which are used to extract plant nutrients and inject toxins that cause abnormal plant growth on many plant species of economic importance (Deinhart, 2011). The literature is replete with reports of pecuniary losses to agriculture, horticulture, forestry and plantation crops due to eriophyid infestations (Ashihara *et al.*, 2004; Debnath and Karmakar, 2013; Van Leeuwen *et al.*, 2010). Likewise, several earlier studies have reported susceptibility of eriophyid mites to a wide range of pesticides (Azevedo *et al.*, 2013; Isaacs *et al.*, 2004) and neem bio-pesticides (Badge *et al.*, 2014). However, studies on eriophyid mite injurious to seabuckthorn are utterly lacking. It is because of the fact that, hitherto, no eriophyid has ever been reported to infest this plant elsewhere. This microscopic mite observed on seabuckthorn caused leaves to become discolored and distorted due to blister development. These blisters dried out leaving behind dead areas on the leaf blade. Presence of multiple blisters on the single leaf due to severe mite infestation greatly reduced the photosynthetic area. Consequently, reduced photosynthetic activity weakened the plant resulting in poor berry production. Keeping in view the enormous industrial utility of this plant and concomitant threat to its scientific cultivation emanating from eriophyid menace, it was prudently thought to devise strategies for its management on

seabuckthorn. Hence, the present manuscript deals with evaluation of some acaricides for the management of this eriophyid mite on seabuckthorn.

MATERIALS AND METHODS

Selection of seabuckthorn species for experimentation

Three species of seabuckthorn viz. *H. rhamnoides*, *H. salicifolia* and *H. tibetana* grow naturally in the North West Himalayan region including cold deserts of Lahaul and Spiti, Himachal Pradesh, India (Rousi, 1971; Zeb, 2004b). Of these, *H. rhamnoides* and *H. salicifolia* are more common and exist as natural stands throughout the two valleys. But, majority of these natural seabuckthorn stands are unmanaged, non-uniform and inaccessible. However, scientifically planned orchards of *H. rhamnoides* were established at a few locations in the Lahaul valley under National Agriculture Innovation Project (NAIP) funded by Indian Council of Agricultural Research (ICAR), New Delhi, India. These sites were easily accessible and had plants of uniform age required for scientific experimentation. Therefore, *H. rhamnoides* was selected for experimentation in the present studies.

Details of the experimental site

The present studies were carried out in village Trilokinath (Altitude: 3200 m amsl, Latitude: 32°40.699' N, Longitude: 76°41.928' E) in the Lahaul valley of Himachal Pradesh, India during summer 2011 and 2012. The selected site had terraced landform where *H. rhamnoides* orchard on about 0.5 ha was established in 2008. The plants were planted at spacing of 3 m within a row and two successive rows were spaced 5 m apart.

Selection of seabuckthorn plants for experimentation

Despite planting at the same time, uniformity lacked in the orchard which could be attributed to some differences in edaphic and agronomic factors inherent in the terraced land. Therefore, relatively uniform plants that measured around 1.5 to 2 m in height and showed symptoms of mite infestation were purposely chosen for assessing the efficacy of applied acaricides in managing eriophyid mite.

Tested acaricides and their doses

An identical dose (0.01%) of four acaricides namely dicofol 18.5EC (0.5 ml/L), propargite 57EC (0.2 ml/L), fenpyroximate 5EC (2.0 ml/L) and hexythiazox 5.45EC (1.8 ml/L) was tested against this blister inducing leaf mite. An unsprayed control was simultaneously maintained to compare the effectiveness of the applied acaricides.

Details of experimentation

The efficacy of acaricides was ascertained on the basis of the number of mites present in 2 mm wide leaf strip incised from center of the leaf specimen before and after application of acaricides as advocated by Pushpa (2006) for coconut perianth mite, *Aceria guerreronis* Keifer with minor modification. Three rows were selected at random from the orchard with each row representing single replication. Fifteen plants (3 plants/treatment/replication) showing symptoms of eriophyid mite infestation as evidenced by presence of blistered leaves were selected. Three infested shoots per plant were aluminum tagged that indicated the applied acaricide. The whole plant

along with tagged shoots was thoroughly sprayed with the test acaricide to prevent probable post spray colonization of leaves on the tagged shoots by mites moving from the unsprayed shoots. Selected acaricides were sprayed twice at ten days interval. Randomization in the acaricide application was achieved by ensuring that plants in the close proximity did not receive sprays of the same acaricide. Two leaves selected at random from each tagged shoot were collected 1 day before spraying (pre-treatment) and 3, 10 days after first and second spray (DAS₁/DAS₂). Thus, eighteen leaves collected from three plants sprayed with same acaricide constituted one replication. While collecting the leaves, care was taken not to hand touch the leaf lamina directly to ensure accuracy in microscopic observations on mite population. The leaves were held in hand from the pedicel, gently plucked and then packed in labeled plastic vials (12cm x 4cm) with perforated caps. Consequently, the vials were brought to the Plant Protection Laboratory of Highland Agricultural Research and Extension Centre, Kukumseri, Lahaul and Spiti of Himachal Pradesh Agricultural University, Palampur, India for recording data.

Data recording and statistical analysis

Data were recorded on the mite population and number of blisters on the leaves, formed as a result of feeding by eriophyid mite. Whereas, all blisters on the single leaf were visually counted, the mite population was microscopically determined by directly examining the leaf specimen under microscope as outlined by Monfreda *et al.* (2010). Care was also taken not to hand touch the leaf lamina while taking observations. For this purpose, single leaf was picked up and held in hand by forceps. Number of blisters on the picked leaf was noted and then a strip measuring 2 mm wide was incised with the help of a small sharp surgical scissor. The incised leaf strip was placed on the glass slide and examined under compound microscope (10X to 40X magnification) for recording mite population. The microscope too had slide adjustment mechanism for hands free observations. The minute and almost transparent moving mites were carefully counted from both sides by reversing the leaf strip with forceps. The data from all the eighteen leaves representing single replication were similarly recorded for all the three replications. The replication wise data were then aggregated and averaged separately to find mean blister count per leaf and mean mite count per 2 mm wide leaf strip for each replication. Subsequently, the data on blister and mite counts were statistically analyzed after square root transformation using CPCS1 software (Cheema and Singh, 1990). Afterwards, per cent change in mite count and blister count at 3, 10 DAS₁/DAS₂ over pre-treatment were also worked out as below:

$$\text{Change in mite count (\%)} = \frac{(MC_n - MC_{ptm})}{MC_{ptm}} \times 100$$

Where, MC_n - mite count on nth day after acaricide application (n = 3, 10); MC_{ptm} - mite count at pre-treatment

$$\text{Change in blister count (\%)} = \frac{(BC_n - BC_{ptm})}{BC_{ptm}} \times 100$$

Where, BC_n - blister count on nth day after acaricide application (n = 3, 10); BC_{ptm} - blister count at pre-treatment.

The per cent changes in mite and blister counts were computed over pre-treatment instead of unsprayed control because changes in the same plants would be an ideal indicator of the efficacy of sprayed acaricides.

RESULTS

Effect of acaricides on population build-up of eriophyid mite

All the four acaricides evaluated against eriophyid mite infesting seabuckthorn effectively checked its spread and prevented blister development on the leaves. It was quite evident from the significant reduction in mite population (Table 1) and blister development (Table 2) on the leaves of sprayed plants as observed at 3 and 10 days after first (DAS₁) and second spray (DAS₂) over pre-treatment. On the other hand, significant increase in mite population and blister count over pre-treatment was observed on the leaves of unsprayed control plants during these times in both the years. At 3DAS₁, maximum 93.8 and 88.9 per cent decrease in mite population over pre-treatment was recorded in plants sprayed with dicofol 18.5EC (0.5 ml/L) in 2011 and 2012, respectively. This corresponded to reduction in mite count from 83.6, 51.2 mites/2mm wide

leaf strip at pre-treatment to 5.2, 5.7 mites/2mm wide leaf strip at 3DAS₁ in two years, respectively. Whereas, the minimum reduction recorded in mite population at 3DAS₁ over pre-treatment was 88.9 per cent in hexythiazox 5.45EC (1.8 ml/L) and 79.1 per cent in propargite 57EC (0.2 ml/L) sprayed plants in 2011 and 2012, respectively. The unsprayed control plants at this time recorded 28.4 and 50.7 per cent surge in mite population over pre-treatment in two years, respectively. Mite population in these unsprayed plants increased from 81.7, 45.4 mites/2mm wide leaf strip at pre-treatment to 104.9, 68.4 mites/2mm wide leaf strip at 3DAS₁ in two respective years (Table 1). The per cent reduction in mite population over pre-treatment was lower at 10DAS₁ when compared to 3DAS₁. In the two years, the highest reduction in mite population over pre-treatment at 10DAS₁ (76.9 %) was observed in hexythiazox 5.45EC (1.8 ml/L) sprayed plants during 2011. On the other hand, the lowest reduction in mite population over pre-treatment at 3DAS₁ was 79.1 per cent in propargite 57EC (0.2 ml/L) sprayed plants during 2012. However, after the second spray of acaricides, mite population decreased and the maximum drop at 3DAS₂ over pre-treatment (98.4, 98.7%) was recorded in fenpyroximate 5EC (2.0 ml/L) and minimum

Table 1: Effect of acaricides on population build-up of eriophyid mite (*Aceria* sp.) on leaves of seabuckthorn

Tested acaricides	Dose (ml/L)	Mean population of eriophyid mite (No./2mm wide leaf strip) ^{1,2,3}					Change in population of eriophyid mite over pre-treatment ⁴ (%)			
		PTM ⁵	3DAS ₁ ⁶	10DAS ₁ ⁶	3DAS ₂ ⁶	10DAS ₂ ⁶	3DAS ₁ ⁶	10DAS ₁ ⁶	3DAS ₂ ⁶	10DAS ₂ ⁶
Dicofol 18.5EC	0.5	83.6 (9.2)	5.2 (2.5) ^a	26.0 (5.2) ^{ab}	3.5 (2.1) ^a	0.5 (1.2) ^a	-93.8	-68.9	-95.8	-99.4
		51.2 (7.2) ^{cd}	5.7 (2.6) ^a	22.4 (4.8) ^a	1.5 (1.6) ^a	0.6 (1.3) ^a	-88.9	-56.3	-97.1	-98.8
Propargite 57EC	0.2	89.8 (9.5)	8.9 (2.8) ^a	23.9 (5.0) ^{ab}	2.8 (1.9) ^a	0.4 (1.2) ^a	-90.1	-73.4	-96.9	-99.6
		39.7 (6.4) ^b	8.3 (3.0) ^{ab}	30.3 (5.6) ^b	1.5 (1.6) ^a	0.2 (1.1) ^a	-79.1	-23.7	-96.2	-99.5
Fenpyroximate 5EC	2.0	95.9 (9.8)	6.3 (2.7) ^a	32.0 (5.7) ^{ab}	1.5 (1.6) ^a	3.7 (1.9) ^a	-93.4	-66.6	-98.4	-96.1
		29.7 (5.5) ^a	5.7 (2.6) ^a	18.3 (4.4) ^a	0.4 (1.2) ^a	0.2 (1.1) ^a	-80.8	-38.4	-98.7	-99.3
Hexythiazox 5.45EC	1.8	93.7 (9.7)	10.4 (3.2) ^a	21.6 (4.7) ^a	7.4 (2.9) ^b	0.5 (1.2) ^a	-88.9	-76.9	-92.1	-99.5
		58.4 (7.7) ^d	10.5 (3.4) ^b	31.6 (5.7) ^b	6.9 (2.8) ^b	9.3 (3.2) ^b	-82.0	-45.9	-88.2	-84.1
Unsprayed control	-	81.7 (9.1)	104.9 (10.3) ^b	140.2 (11.9) ^c	198.3 (14.1) ^c	238.3 (15.5) ^b	+28.4	+71.6	+142.7	+246.8
CD (0.05)		NS (0.7)	68.4 (8.3) ^c (1.6) (0.6)	90.6 (9.6) ^c (0.8) (0.6)	(0.5) (0.6)	(1.2) (0.8)	+50.7	+99.6	+142.5	+233.5

¹Data in the first and second lines pertain to years 2011 and 2012, respectively; ²Figures in the parentheses are square root transformed means; ³Figures in the parentheses followed by the same alphabet are non-significant with each other; ⁴Negative (-) and positive (+) signs denote decrease and increase in mite population, respectively; ⁵Pre-treatment; ⁶Days after first and second spray

Table 2: Effect of acaricides on development of blisters due to feeding by eriophyid mite (*Aceria* sp.) on leaves of seabuckthorn

Tested acaricides	Dose (ml/L)	Mean blister count (No./leaf) ^{1,2,3}					Change in blister count over pre-treatment ⁴ (%)			
		PTM ⁵	3DAS ₁ ⁶	10DAS ₁ ⁶	3DAS ₂ ⁶	10DAS ₂ ⁶	3DAS ₁ ⁶	10DAS ₁ ⁶	3DAS ₂ ⁶	10DAS ₂ ⁶
Dicofol 18.5EC	0.5	1.9 (1.7)	1.8 (1.7)	1.5 (1.6)	1.3 (1.5) ^a	0.7 (1.3) ^a	-5.3	-21.1	-31.6	-63.2
		1.4 (1.5)	1.3 (1.5)	1.1 (1.4) ^a	0.6 (1.3) ^a	0.3 (1.1) ^a	-7.1	-21.4	-57.1	-78.6
Propargite 57EC	0.2	2.1 (1.8)	2.0 (1.7)	1.6 (1.6)	1.3 (1.5) ^a	0.4 (1.2) ^a	-4.8	-23.8	-38.1	-81.0
		1.5 (1.6)	1.4 (1.5)	1.1 (1.4) ^a	0.7 (1.3) ^a	0.5 (1.2) ^a	-6.7	-26.7	-53.3	-66.7
Fenpyroximate 5EC	2.0	2.0 (1.7)	1.8 (1.7)	1.6 (1.6)	1.0 (1.4) ^a	0.6 (1.3) ^a	-10.0	-20.0	-50.0	-70.0
		1.6 (1.6)	1.5 (1.6)	1.1 (1.4) ^a	0.5 (1.2) ^a	0.5 (1.2) ^a	-6.3	-31.3	-68.8	-68.8
Hexythiazox 5.45EC	1.8	2.0 (1.7)	1.8 (1.7)	1.6 (1.6)	1.2 (1.5) ^a	0.6 (1.3) ^a	-10.0	-20.0	-40.0	-70.0
		1.6 (1.6)	1.5 (1.6)	1.2 (1.5) ^a	0.6 (1.3) ^a	0.7 (1.3) ^a	-6.3	-25.0	-62.5	-56.3
Unsprayed control	-	1.7 (1.6)	1.8 (1.7)	1.9 (1.7)	2.6 (1.9) ^b	3.5 (2.1) ^b	+5.9	+11.8	+52.9	+105.9
CD (0.05)		1.2 (1.5) NS	1.4 (1.6) NS	2.0 (1.7) ^b NS (0.1)	2.1 (1.8) ^b (0.2) (0.2)	2.4 (1.8) ^b (0.2)	+16.7	+66.7	+75.0	+100.0

¹Data in the first and second lines pertain to years 2011 and 2012, respectively; ²Figures in the parentheses are square root transformed means; ³Figures in the parentheses followed by the same alphabet are non-significant with each other; ⁴Negative (-) and positive (+) signs denote decrease and increase in blister count, respectively; ⁵Pre-treatment; ⁶Days after first and second spray.

(92.1, 88.2%) in hexythiazox 5.45EC (1.8 ml/L) sprayed plants in two years, respectively. The plants sprayed with fenpyroximate 5EC (2.0 ml/L) had only 1.5, 0.4 mites/2mm wide leaf strip at 3DAS₂ as compared to 95.9, 29.7 mites/2mm wide leaf strip at pre-treatment in 2011 and 2012, respectively. The corresponding figures in plants sprayed with hexythiazox 5.45EC (1.8 ml/L) were 7.4, 6.9 mites/2mm wide leaf strip at 3DAS₂ and 93.7, 58.4 mites/2mm wide leaf strip at pre-treatment in two years, respectively. At this time interval, mite population on unsprayed control plants increased by more than 142 per cent over pre-treatment in both the years. The respective mite count for two years in these unsprayed control plants was 198.3, 110.1 mites/2mm wide leaf strip at 3DAS₂ as against 81.7, 45.4 mites/2mm wide leaf strip at pre-treatment (Table 1). At 10DAS₂, all the acaricides recorded more than 96 per cent reduction in mite population over pre-treatment in both the years except hexythiazox 5.45EC (1.8 ml/L) sprayed plants in 2012 (84.1%). The mean mite count declined below 1.0 mite/2mm wide leaf strip in both the years except 9.3 mites/2mm wide leaf strip recorded in hexythiazox 5.45EC (1.8 ml/L) sprayed plants in 2012. Whereas, the control plant during this time recorded 246.8 and 233.5 per cent more mite population when compared to pre-treatment and it corresponded to 238.3, 151.4 mites/2mm wide leaf strip at 10DAS₂ as compared to 81.7, 45.4 mites/2mm wide leaf strip at pre-treatment in respective years (Table 1).

Effect of acaricides on development of blisters on seabuckthorn leaves

In 2011, plants sprayed with evaluated acaricides reduced blister count on leaves by 20.0 to 23.8 per cent as recorded at 10DAS₁ in comparison to pre-treatment although all were statistically non-significant to each other. However, 11.8 per cent increase in blister count over pre-treatment was observed in unsprayed control plants during the same time. The leaves of these plants recorded increase in the mean blister count from 1.7 blisters/leaf at pre-treatment to 1.9 blisters/leaf at 10DAS₁. During 2012 also, tested acaricides did not differ significantly in reducing the blister count at 10DAS₁. Nevertheless, the blister count on plants sprayed with acaricides varied from 1.1 to 1.6 blisters/leaf and was significantly lower than the unsprayed control plants which recorded 1.9, 2.0 blisters/leaf in two years, respectively. The highest reduction in blister count (31.3%) over pre-treatment during this time was recorded in those plants that were sprayed with fenpyroximate 5EC (2.0 ml/L) and the lowest (21.4%) in dicofol 18.5EC (0.5 ml/L) sprayed plants in this year. Correspondingly, the mean numbers of blisters per leaf decreased from 1.6, 1.4 at pre-treatment in respective acaricides to 1.1 at 10DAS₁ in both the acaricides (Table 2). On the other hand, the control plants recorded 66.7 per cent increase in blister count over pre-treatment at this time in the same year which corresponded to 1.2 and 2.0 blisters/leaf at pre-treatment and 10DAS₁, respectively. Similarly at 10 DAS₂, all the acaricides resulted in statistically similar reductions in blister count. However, the plants sprayed with propargite 57EC (0.2 ml/L) recorded the highest (81.0%) and dicofol 18.5EC (0.5 ml/L) the lowest (63.2%) reduction over pre-treatment in 2011. During 2012, dicofol 18.5EC (0.5 ml/L) sprayed plants again recorded the highest reduction in blister

count over pre-treatment (78.6%) but the lowest (56.3%) was recorded in plants sprayed with hexythiazox 5.45EC (1.8 ml/L). Propargite 57EC (0.2 ml/L) sprayed plants during this year registered 66.7 per cent reduction in blister count over pre-treatment. On the other hand, blister count in the unsprayed control plants increased by 105.9 and 100 per cent over pre-treatment at 10DAS₂ in 2011 and 2012, respectively. The mean blister count on these unsprayed plants increased from 1.7, 1.2 blisters/leaf at pre-treatment to 3.5, 2.4 blisters/leaf at 10DAS₂ in two years, respectively (Table 2).

DISCUSSION

The eriophyid mite infestations on seabuckthorn have not yet been documented in the published literature. Whereas, wealth of literature exists on evaluation of numerous acaricides including dicofol 18.5EC, propargite 57EC, fenpyroximate 5EC and hexythiazox 5.45EC against many mites infesting various agriculture, horticulture and plantation crops. Therefore, the efficacy of acaricides used in present investigations has been discussed vis-à-vis earlier studies, albeit against mites including eriophyids damaging other crops. For instance, Kumar *et al.*, (2009) reported highest reduction (74.2%) in *Tetranychus urticae* Koch population on brinjal plants sprayed with dicofol 18.5EC (4.0 ml/L) followed by fenpyroximate 5EC (0.8 ml/L) (66.3%) and propargite 57EC (4.0 ml/L) (66.1%). On the other hand, Murthy and Bhushan (2010) found fenpyroximate 5EC (500-625 ml/ha) to be more effective in controlling eriophyid mite, *Aceria lycopersici* Solff on tomato and tetranychid mite, *Tetranychus cucurbitae* Rahman and Sapa on bottle gourd as compared to dicofol 18.5EC (1375 ml/ha). Reddy and Latha (2013) conducted field experiments to test the bio-efficacy of acaricides against two spotted spider mite, *T. urticae* on ridge gourd and found that hexythiazox 5.4EC (25 g a.i./ha) and fenpyroximate 5EC (30 g a.i./ha) were comparatively more efficacious and caused 66.5-70.6 and 69.9-82.8 per cent mite mortality at 14 days after spray (DAS) as compared to dicofol 18.5EC (250 g a.i./ha) (22.9-34.3%) and propargite 57EC (612 g a.i./ha) (36.3-55.6%). In the same way, Chakrabarti and Sarkar (2014) reported that fenpyroximate 5EC (60 g a.i./ha) and propargite 57EC (1000 g a.i./ha), each sprayed twice, gave 68.2 and 72.2 per cent control of *Polyphagotarsonemus latus* (Banks) infesting chilli. Likewise, Khajuria and Sharma (2010) reported moderate toxicity of propargite 57EC (0.057 %) against the two spotted spider mite and found it to be safer to the predatory mites. On the other hand, Kavya *et al.* (2015) concluded that fenpyroximate 5EC was more toxic to adult females of *T. urticae* with LC₅₀ value of 1.91 ppm than dicofol 18.5EC (27.84 ppm) and propargite 57EC (31.73 ppm). A careful perusal of this literature would reveal that these acaricides had proven effective in managing eriophyid and tetranychid mites. Still, the variations in efficacy of same acaricide apparent in these studies could be attributed to varying doses and different target mite species besides diverse agro-ecological situations. However, the high efficacy of all these acaricides against eriophyid on seabuckthorn could be ascribed to its earlier non-exposure to any acaricide that probably made its populations highly susceptible to the applied acaricides. Villavicencio *et al.* (2014) found fenpyroximate 5EC effective for preventive and curative control

of eriophyid mite, *Aceria aloinis* Keifer infesting aloe. These authors noted that ornamental aloe plants sprayed with fenpyroximate 5EC (16 fl oz/100 gallons water) had lower damage severity than the inoculated controls and no mites were found in the plants even one year after acaricide application. Their findings further revealed that curative control of infested plants was effective when plant tissue damaged by aloe mites was removed before acaricide application. Likewise, Koehler *et al.* (1987) observed that though dicofol 18.5EC (0.4 pounds a.i./100 gallons water) and propargite 57EC (0.3 pounds a.i./100 gallons water) suppressed fuchsia gall mite, *Aculops fuchsiae* Keifer, but the degree of suppression was insufficient for lasting control because of rapid mite reproduction. In the present studies too, though the applied acaricides suppressed the eriophyid mite population initially as observed at 3DAS₁, but it increased slightly afterwards as revealed by lesser reductions over pre-treatment at 10DAS₁ as compared to at 3DAS₁. Nevertheless, the second spray of these acaricides effectively controlled the eriophyid mite as shown by reductions in its population (84.1-99.6%) and blister count (56.3-81.0%) on the leaves of seabuckthorn at 10DAS₂. The efficacy of the acaricides could probably be improved if misshapen blistered leaves having colonies of eriophyid mites were removed prior to their application, as was observed by Villavicencio *et al.* (2014) in case of *A. aloinis* infesting ornamental aloe. This improved effectiveness of acaricides against *A. aloinis* could have resulted from lower mite inoculum subsequent to removal of shoots damaged by mites. However, manual removal of blistered seabuckthorn leaves would be practicable and justifiable only when these were few signifying low level of eriophyid infestation. Singh and Singh (2014) reported that fenpyroximate 5EC (1.0 ml/L), propargite 57EC (2.0 ml/L) and dicofol 18.5EC (2.5 ml/L) caused 81.3, 70.9 and 56.1 per cent mean mortality of tetranychid mite, *Tetranychus neocaledonicus* Andre infesting brinjal, respectively. The corresponding reductions in mite population over control were 79.1, 68.4 and 52.2 per cent, respectively. Similarly, the mean population of *T. urticae* infesting okra was reduced by 77.3, 71.9 and 66.9 per cent over control in plots sprayed with fenpyroximate 5EC (0.5 ml/L), propargite 57EC (2.0 ml/L) and dicofol 18.5EC (2.0 ml/L), respectively (Singh *et al.*, 2014). These studies lent support to present results where fenpyroximate 5EC (2.0 ml/L), propargite 57EC (0.2 ml/L) and dicofol 18.5EC (0.5 ml/L) caused significant eriophyid mortalities and reduced its population by 96.1, 99.6, 99.4 per cent over pre-treatment in 2011 and 99.3, 99.5, 98.8 per cent in 2012, respectively. Likewise, Reddy *et al.* (2014) also observed decrease in mortality of *T. urticae* on cucumber from 74.0, 81.3 per cent at 1DAS to 22.3, 39.2 per cent at 14DAS in dicofol 18.5EC (2.7 ml/L) and propargite 57EC (2.0 ml/L) sprayed plants, respectively. The corresponding mortalities recorded in fenpyroximate 5EC and hexythiazox 5.45EC both sprayed at the rate of 1.25 ml/L were 79.2, 83.9 per cent at 1DAS and 59.7, 55.7 per cent at 14DAS, respectively. The observations of these authors also corroborated the outcomes of present studies where reduction in eriophyid mite population decreased from 93.8, 90.1, 93.4, 88.9 per cent at 3DAS₁ to 68.9, 73.4, 66.6, 76.9 per cent at 10DAS₁ in 2011 and from 88.9, 79.1, 80.8, 82.0 per cent at 3DAS₁ to 56.3, 23.7, 38.4, 45.9 per cent at 10DAS₁ in 2012 in

case of dicofol 18.5EC (0.5 ml/L), propargite 57EC (0.2 ml/L), fenpyroximate 5EC (2.0 ml/L) and hexythiazox 5.45EC (1.8 ml/L) sprayed plants, respectively. The increase in population a few days after first acaricide spray possibly occurred because of rapid multiplication due to short life cycle of the eriophyid mite. Present findings are also supported by Sood *et al.* (2015) who found dicofol 18.5EC (250 g a.i./ha) and propargite 57EC (430 g a.i./ha) effective against *T. urticae* on cucumber and observed reductions in its mean population from 13.7, 14.6 and 101.7, 111.3 mites/5 leaves at pre-treatment to 3.6, 3.9 and 1.6, 2.3 mites/5 leaves at 21 DAS₂ in autumn and summer crops, respectively grown under protected environment. Field bio-efficacy of fenpyroximate 5EC (15, 20, 25 g a.i./ha) and dicofol 18.5EC (250 g a.i./ha) against red spider mite, *Tetranychus cinnabarinus* (Boisd.) infesting brinjal showed that these acaricides effectively controlled this mite (Naik *et al.*, 2006) and both these acaricides were found quite effective in current investigations as well. However, the findings of Forti *et al.* (1994) suggested fenpyroximate 5EC to be very effective against tetranychid, *Panonychus ulmi* (Koch.) and comparatively less effective against the eriophyid *Aculus schlechtendali* (Nal.) infesting apple (cv-Golden Delicious) in Italy. Thus, their results were in contradiction to the present results where fenpyroximate 5EC (2.0 ml/L) conclusively proved highly effective against seabuckthorn eriophyid and caused more than 96 per cent reduction in mite population and around 70 per cent reduction in blister development on leaves at 10DAS₂. The probable reasons for the lower efficacy of this acaricide against the eriophyid *A. schlechtendali* could be inherent differential susceptibility of the two species infesting different hosts. It could also be possible that the apple eriophyid had developed some level of resistance to the sprayed acaricide due to relatively heavy pesticides application usually resorted to in managed apple orchards. On the other hand, eriophyids on seabuckthorn were never subjected to pesticide pressure earlier and perhaps, therefore, comprised of highly susceptible population that readily succumbed to acaricide application. This inference invariably underscores the utility of rotational application of acaricides in management strategies against eriophyid mite menace. Moreover, some earlier studies had also indicated its comparative safety to natural enemies of mites. For instance, Lima *et al.* (2013) found fenpyroximate 5EC not only promising against coconut mite, *Aceria guerreronis* Keifer but also safer to its predator *Neoseiulus baraki* Athias-Henriot, as it recorded higher LC₅₀ and LC₉₀ values in laboratory studies. Similarly, Kim and Paik (1996) evaluated relative toxicity of fenpyroximate 5EC to kanzawa spider mite, *Tetranychus kanzawai* Kishida, an arthropod pest of tea and predatory mite, *Amblyseius womersleyi* Schicha under laboratory conditions and found it to be less toxic to adult females and eggs of *A. womersleyi* than *T. kanzawai*. They also reported no significant reduction in reproduction and hatchability of *A. womersleyi* eggs at concentrations of 6.25 to 50 ppm. Also, Bala *et al.* (2015) found very low population (1.4 mites/cm²) of eriophyid mite, *Aceria tulipae* (Keifer) infesting garlic at 10DAS in dicofol 18.5EC (2.5 ml/L) sprayed plots and concluded that it was most effective acaricide against this eriophyid. Their studies also confirmed the results of present investigations where dicofol 18.5EC (0.5 ml/L) sprayed plants had just 3.5, 1.5 and 0.5, 0.6 mites/2mm wide

leaf strip at 3DAS₂ and 10DAS₂ in two years, respectively.

In corollary, it could be concluded that evaluated acaricides effectively countered the population build-up of eriophyid mite on seabuckthorn. The blister formation on the leaves due to mite feeding was also substantially reduced in plants that received acaricide application. Hence, these acaricides could be used to manage eriophyid mite on seabuckthorn.

ACKNOWLEDGEMENT

The authors gratefully acknowledge and express gratitude to the Indian Council of Agricultural Research (ICAR), New Delhi for providing financial support under National Agriculture Innovation Project (NAIP) to carry out these research investigations.

REFERENCES

- Abid, H., Hussain, A. and Ali, S. 2007. Physicochemical characteristics and fatty acid composition of seabuckthorn (*Hippophae rhamnoides* L.) oil. *J. Chemical Society of Pakistan*. **29**: 256-259.
- Ashihara, W., Kondo, A., Shibao, M., Tanaka, H., Hiehata, K. and Izumi, K. 2004. Ecology and control of eriophyid mites injurious to fruit trees in Japan. *Japan Agricultural Research Quarterly*. **38**(1): 31-41.
- Azevedo, L. H., Moraes, G. J., Yamamoto, P. T. and Zanardi, O. Z. 2013. Development of a methodology and evaluation of pesticides against *Aceria litchii* and its predator *Phytoseius intermedius* (Acari: Eriophyidae, Phytoseiidae). *J. Economic Entomology*. **16**: 2183-2189.
- Bagde, A. S., Patil, P. D. and Pashte, V. V. 2014. Studies on efficacy of neem bio-pesticides against eriophyid mite (*Aceria guerreronis* Keifer.). *The Bioscan*. **9**(1): 341-346.
- Bala, S. C., Karmakar, K. and Ghosh, S. K. 2015. Population dynamics of mite, *Aceria tulipae* (Keif.) on garlic (*Allium sativum* L.) and its management under Bengal Basin. *International J. Science Environment and Technology*. **4**(5): 1365-1372.
- Catherine, U., Dilys, B., Dawn, C., Samantha, C., Nicole, G., Richard, I., Erica, R., Jill, M. G. S., Tera, S., Shaina, T. C., Regina, C. W. and Sara, Z. 2012. Seabuckthorn (*Hippophae rhamnoides*): An evidence based systematic review by the Natural Standard Research Collaboration. *Alternative and Complementary Therapies*. **18**(4): 207-219.
- Chakrabarti, S. and Sarkar, P. K. 2014. Studies on efficacy of some acaricidal molecules for the management of *Polyphagotarsonemus latus* (Banks) (Acari: Tarsonemidae) infesting chilli (*Capsicum annum* L.) in West Bengal. *Current Biotica*. **7**(4): 299-305.
- Chaman S., Syed, N. H., Danish, Z. and Khan, F. Z. 2011. Phytochemical analysis, antioxidant and antibacterial effects of seabuckthorn berries. *Pakistan J. Pharmaceutical Sciences*. **24**(3): 345-351.
- Chauhan, A. S., Negi, P. S. and Ramteke, R. S. 2007. Antioxidant and antibacterial activities of aqueous extract of seabuckthorn (*Hippophae rhamnoides*) seeds. *Fitoterapia*. **78**: 590-592.
- Cheema, H. S. and Singh, B. 1990. CPCS1: Computer programme package for the analysis of commonly used experimental designs. *Punjab Agricultural University, Ludhiana, Punjab*.
- Debnath, P. and Karmakar, K. 2013. Garlic mite, *Aceria tulipae* (Keifer) (Acari: Eriophyoidea) - a threat for garlic in West Bengal, India. *International J. Acarology*. **30**(2): 89-96.
- Deinhart, N. 2011. Tiny monsters: *Aceria aloinis*. *Cactus and Succulent J*. **83**: 120-122.
- Dhyani, D., Maikhuri, R. K. and Dhyani, S. 2011. Utilization pattern of seabuckthorn in food and health care system in central Himalaya Uttarakhand. In: Emerging Trends in R & D on Health Protection and Environmental Conservation, Proceedings of National Conference on Seabuckthorn, 1-3 December, 2011, CSKHPKV Palampur, India. pp 270-278.
- Fan, J., Ding, X. and Gu, W. 2007. Radical-scavenging proanthocyanidins from seabuckthorn seed. *Food Chemistry*. **102**(1): 168-177.
- Forti, D., Loriatti, C., Angeli, G. and Catoni, M. 1994. Two new acaricides, pyridaben and fenpyroximate: evaluation of their efficacy on *Panonychus ulmi* (Koch.) and *Aculus schlechtendali* (Nal.) and their non-target effects on beneficial arthropods. *Informatore Fitopatologico*. **44**(7-8): 38-42.
- Geetha, S., Ram, M. S. and Singh, V. 2002. Anti-oxidant and immunomodulatory properties of seabuckthorn (*Hippophae rhamnoides*)-an in-vitro study. *J. Ethnopharmacology*. **79**(3): 373-378.
- Hibasami, H., Mitani, A., Katsuzaki, H., Imai, K., Yoshioka, K. and Komiya, T. 2005. Isolation of five types of flavonol from seabuckthorn (*Hippophae rhamnoides*) and induction of apoptosis by some of the flavonols in human promyelotic leukemia HL-60 cells. *International J. Molecular Medicine*. **15**: 805-809.
- Isaacs, R., Morrone, V. and Gajek, D. 2004. Potential acaricides for management of blueberry bud mite in Michigan blueberries. *HortTechnology*. **14**(2): 188-191.
- Kallio, H., Yang, B., Peippo, P., Tahvonen, R. and Pan, R. 2004. Triacylglycerols, glycerophospholipids, tocopherols and tocotrienols in berries and seeds of two subspecies (ssp. *sinensis* and *mongolica*) of seabuckthorn (*Hippophae rhamnoides*). *J. Agricultural and Food Chemistry*. **50**: 3004-3009.
- Kavya, M. K., Srinivasa, N., Ravi, G. B. and Vidyashree, A. S. 2015. Relative toxicity of selected acaricides on two spotted spider mite (*Tetranychus urticae*) of brinjal. *The Bioscan*. **10**(2): 605-608.
- Khajuria, D. R. and Sharma, J. P. 2010. Bio-efficacy of various acaricides against the phytophagous mites on apple and their sensitivity to the phytoseiid mites. *Pest Management and Economic Zoology*. **180**(1/2): 116-121.
- Kim, S. S. and Paik, C. H. 1996. Comparative toxicity of fenpyroximate to the predatory mite *Amblyseius womersleyi* Schicha and the kanzawa spider mite, *Tetranychus kanzawai* Kishida (Acarina: Phytoseiidae, Tetranychidae). *Applied Entomology and Zoology*. **31**(3): 369-377.
- Koehler, C. S., Allen, W. W. and Costello, L. R. 1987. Fuchsia gall mite management. *Ornamentals Northwest Archives*. **10**(3): vii-viii.
- Kumar, S. V., Chinnah, C., Muthah, C. and Sadasakthi, A. 2009. Field evaluation of certain acaricide/insecticide molecules for their bio-efficacy against *Tetranychus urticae* Koch on brinjal. *Karnataka J. Agricultural Sciences*. **22**(3): 705-706.
- Li, T. S. C., Beveridge, T. H. J. and Drover, J. C. G. 2007. Phytosterol content of seabuckthorn (*Hippophae rhamnoides* L.) seed oil: Extraction and identification. *Food Chemistry*. **01**: 1633-1639.
- Lima, D. B., Monteiro, V. B., Guedes, R. N. C., Siqueira, H. A. A., Pallini, A. and Gondim Jr., M. G. C. 2013. Acaricide toxicity and synergism of fenpyroximate to coconut mite predator *Neoseiulus baraki*. *Biocontrol*. **58**(5): 595-605.
- Luhua, Z., Ying, T., Zhengyu, Z. and Guangji, W. 2004. Determination of alpha-tocopherol in the traditional Chinese medicinal preparation seabuckthorn oil capsule by non-aqueous reversed phase-HPLC. *Chemical and Pharmaceutical Bulletin*. **52**: 150-152.
- Monfreda, R., Lekveishvili, M., Petanovic, R. and Amrine Jr., J. W. 2010. Collection and detection of eriophyid mites. *Experimental and Applied Acarology*. **51**: 273-282.

- Murthy, K. S. R. K. and Bhushan, S. V. 2010.** Bio-efficacy of fenpyroximate 5% EC for the effective control of eriophyid mite (*Aceria lycopersici* Solff.) on tomato (*Lycopersicon esculentus* L.) and tetranychid mite, *Tetranychus cucurbitae* on bottle gourd (*Lagenaria* sp.) *Pestology*. **34(1)**: 36-40.
- Naik, R. L., Lolage, G. R., Kale, V. D. and Dethle, M. D. 2006.** Field bio-efficacy of flufenzin and fenpyroximate on red spider mite, *Tetranychus cinnabarinus* (Boisd.) infesting brinjal. *J. Entomological Research*. **30(2)**: 133-137.
- Negi, P. S., Chauhan, A. S., Sadia, G. A., Rohinishree, Y. S. and Ramteke, R. S. 2005.** Antioxidant and antibacterial activities of various seabuckthorn (*Hippophae rhamnoides* L.) seed extracts. *Food Chemistry*. **92**: 119-124.
- Pinteau, A., Marpeau, A., Faye, M., Socaciu, C. and Gleizes, M. 2001.** Polar lipid and fatty acid distribution in carotenolipoprotein complexes extracted from seabuckthorn fruits. *Phytochemical Analysis*. **12(5)**: 293-298.
- Pushpa, V. 2006.** Management of coconut perianth mite, *Aceria guerreronis* Keifer. M. Sc. (Agri.) thesis submitted to University of Agricultural Sciences, Dharwad, Karnataka, India.
- Reddy, D. S. and Latha, M. P. 2013.** Efficacy of certain new acaricides against two spotted spider mite, *Tetranychus urticae* Koch. on ridge gourd. *Pest Management in Horticultural Ecosystems*. **19(2)**: 199-202.
- Reddy, D. S., Nagaraj, R., Latha, M. P. and Chowdary, R. 2014.** Comparative evaluation of novel acaricides against two spotted spider mite, *Tetranychus urticae* Koch. infesting cucumber (*Cucumis sativus*) under laboratory and green house conditions. *The Bioscan*. **9(3)**: 1001-1005.
- Rousi, A. 1971.** The genus *Hippophae* L. - a taxonomic study. *Annales Botanici Fennici*. **8**: 177-227.
- Sabir, S. M., Maqsood, H., Ahmed, S. D., Shah, A. H. and Khan, M. Q. 2005.** Chemical and nutritional constituents of seabuckthorn (*Hippophae rhamnoides* ssp. *turkestanica*) berries from Pakistan. *Italian J. Food Science*. **17(4)**: 455-462.
- Sharma, S. K., Sharma, P. C. and Kumar, P. 2011.** Status of insect-pests of seabuckthorn and their nature of damage in Lahaul valley of Himachal Pradesh. In: Emerging Trends in R & D on Health Protection and Environmental Conservation, Proceedings of National Conference on Seabuckthorn, 1-3 December, 2011, CSKHPKV Palampur, India. pp. 32-36.
- Singh, A., Samant, S. S., Lal, M., Sharma, P., Butola, J. S. and Marpa, S. 2011.** Assessment, mapping and harnessing economic potential of *Hippophae* species for the socio-economic development of tribal communities in Himachal Pradesh, India. In: Emerging Trends in R & D on Health Protection and Environmental Conservation, Proceedings of National Conference on Seabuckthorn, 1-3 December, 2011, CSKHPKV Palampur, India. pp. 260-269.
- Singh, P. and Singh, R. N. 2014.** Interactions of abiotic factors with *Tetranychus neocaledonicus* Andre and its management by newer acaricides in brinjal ecosystem. *The Ecoscan*. **VI**: 355-359.
- Singh, S. K., Singh, A. P. and Singh, R. N. 2014.** Comparative bio-efficacy of bio-pesticides and new molecules of acaricides in the management of *Tetranychus urticae* Koch (Acari: Tetranychidae) in okra. *The Ecoscan*. Special issue. **VI**: 279-283.
- Singh, V. 2003.** Geographical adaptation and distribution of seabuckthorn resources. In: (Editor-in-chief) Singh, V., Seabuckthorn- a multipurpose wonder plant, Vol. I. Daya Publishing House, New Delhi. pp. 21-34.
- Sood, A. K., Sood, S. and Singh, V. 2015.** Efficacy evaluation of spiromesifen against red spider mite, *Tetranychus urticae* Koch on parthenocarpic cucumber under protected environment. *The Bioscan*. **10(3)**: 963-966.
- Suleyman, H., Gumustekin, K. and Taysi, S. 2002.** Beneficial effects of *Hippophae rhamnoides* L., on nicotine induced oxidative stress in rat blood compared with vitamin E. *Biological and Pharmaceutical Bulletin*. **25(9)**: 1133-1136.
- Van Leeuwen, T., Witters, J., Nauen, R., Duso, C. and Tirry, L. 2010.** The control of eriophyid mites: state of the art and future challenges. *Experimental and Applied Acarology*. **51**: 205-224.
- Villavicencio, L. E., Bethke, J. A., Dahlke, B. Mey, B. V. and Corkidi, L. 2014.** Curative and preventive control of *Aceria aloinis* (Acari: Eriophyidae) in Southern California. *J. Economic Entomology*. **107(6)**: 2088-2094.
- Wang, B., Lin, L., Ni, Q. and Su, C. 2011.** *Hippophae rhamnoides* Linn. for treatment of diabetes mellitus: A review. *J. Medicinal Plants Research*. **5(13)**: 2599-2607.
- Weller, P. and Breithaupt, D. E. 2003.** Identification and quantification of zeaxanthin esters in plants using liquid chromatography-mass spectrometry. *J. Agricultural and Food Chemistry*. **51**: 7044-7049.
- Zeb, A. 2004a.** Chemical and nutritional constituents of seabuckthorn juice. *Pakistan J. Nutrition*. **3**: 99-106.
- Zeb, A. 2004b.** Important therapeutic uses of seabuckthorn (*Hippophae*): a review. *J. Biological Sciences*. **4**: 687-693.

NATIONAL ENVIRONMENTALISTS ASSOCIATION AND ITS OFFICIAL ORGAN



The Bioscan

An International Quarterly Journal of Life Science

Started in 1988, the National Environmentalists Association has been reorganized in 2006 and now is an association functioning with full vigour and new impetus to meet its objectives with the co-operation of like minded environment conscious academicians from different parts of the nation.

MEMBERSHIP OF THE ASSOCIATION

Any graduate having interest in environmental conservation and protection of nature and natural resources can be the member of the association.

To be the member of the association the application form given below should be duly filled up and sent to the Secretary of the association along with a demand draft of Rs. 750/- (After the 25% concession) for annual membership and Rs. 7500/- (After the 25% concession) for life membership.

FELLOWSHIP OF THE ASSOCIATION

The Association is awarding FELLOWSHIP to deserving academicians / researchers /scientists who are LIFE MEMBERS of the Association after reviewing their bio-data by the Fellows and the Executive Members of the association. The Fellows are privileged to write **F.N.E.A.** after their names .The prestigious Fellowship also includes a citation in recognition of their contribution to society in general and the endeavour for the noble cause of environment in particular.

AWARDS OF THE ASSOCIATION

The Association in its Seminars and Conferences provides the following category of awards on annual basis.

1. **The young scientists award** : It is given to the researchers below the age of 35 years.
2. **The senior scientists award** : It is awarded to the academicians above the age of 35 years.

3. **The best paper award**: It is awarded to the contributor of the Journal **The Bioscan** during the year.
4. **The best paper presentation award** : It is awarded to the scholar whose presentation is the best other than the young scientist category.
5. **The best oration award** : It is awarded to the scholar who delivered invited speech.
6. **The recognition award** : It is awarded to those senior scholars who have contributed to the subject through their continued research .
7. **The environmental awareness award** : It is awarded to those who, apart from their research contribution, have done commendable extension work for environmental betterment.

The number of recipients of award in each category will vary depending upon the recommendation of the panel of judges and the executive committee. The association has the provision to institute awards in the name of persons for whom a with desired sum is donated in consultation with the executive body.

PUBLICATION OF THE ASSOCIATION

In order to provide a platform to a vast group of researchers to express their views and finding of research as well as to promote the attitude of quality research among the scholars of younger generation the association publishes an international quarterly journal – **THE BIOSCAN (ISSN:0973-7049)**. For the benefit of the potential contributors **instructions to authors** is given separately in this journal. However, the details regarding the journal and also the association can be seen on our website www.thebioscan.in.

Cont. P. 078